

Analysis of SANS and USANS Data

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NCNR Summer School

Neutron Small Angle Scattering
and Reflectometry

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Outline

- Basic Equations
- Model-independent methods
 - Guinier, Porod, Invariant
- Non-Linear Model Fitting
 - Particles, Polymers, Materials
- Global Fitting
- Anisotropic Scattering
- Transforms
- Ab initio modeling

Easy



More involved



The Basics

Starting from:
$$\frac{d\sigma}{d\Omega}(\vec{q}) = \frac{1}{N} \left| \sum_i^N b_i e^{i\vec{q}\cdot\vec{r}} \right|^2$$

- We can replace the sum over atoms with an integral over the scattering length density

$$\sum_i^N b_i \rightarrow \int_V \rho(\vec{r}) d\vec{r}$$

- Normalizing by sample volume and introducing scattering length density

$$\frac{d\Sigma}{d\Omega}(\vec{q}) = \frac{N}{V} \frac{d\sigma}{d\Omega}(\vec{q}) = \frac{1}{V} \left| \int_V \rho(\vec{r}) e^{i\vec{q}\cdot\vec{r}} d\vec{r} \right|^2$$

- Inhomogeneities in $\rho(\vec{r})$ give rise to small angle scattering

$\Sigma = \sigma/V$ is the “macroscopic cross section”



Scattering Basis

Different systems each have a natural basis - and all are equivalent

- This is especially true if the scattering is from “countable” units

$$\left| \int_V f(\vec{r}) d\vec{r} \right|^2 \rightarrow \sum_i^N \sum_j^N f(\vec{r}_i - \vec{r}_j)$$

Polymers

monomer unit

Particulates

per particle

Proteins

polypeptide subunits

- A statistical description may also be appropriate

$$\rho(r) \rightarrow \gamma(r)$$

Non-particulate

correlation function



Measured Intensity

After reducing the raw data - you will typically have:

$I(Q)$ vs. Q

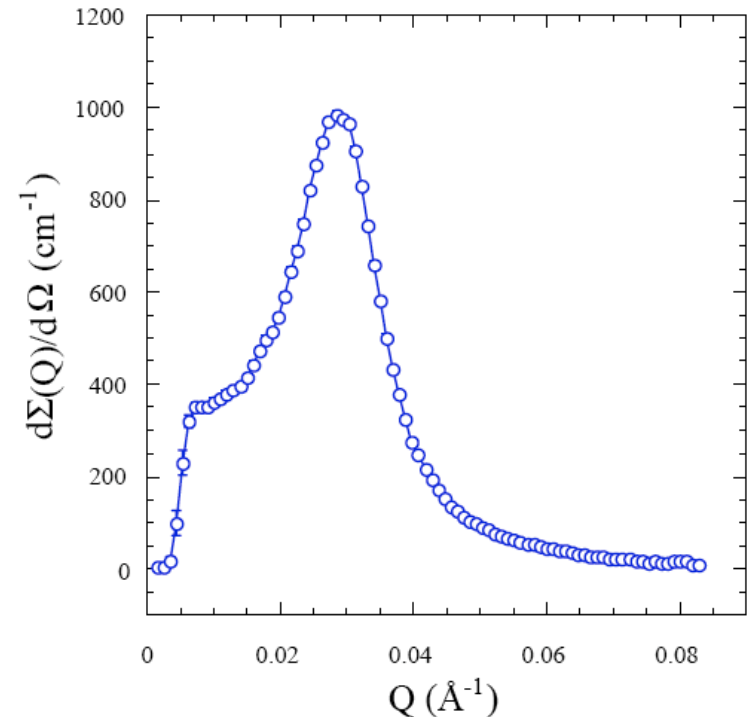
Units of intensity are $\text{cm}^{-1} \text{ster}^{-1}$

Units of Q are length^{-1}

$$I(Q) = \frac{8\pi\xi^3 (\Delta\rho)^2 \phi(1-\phi)}{(1 + (Q\xi)^2)^2}$$

$$I(Q) = \frac{\phi}{V_p} \left[\frac{3V_p (\Delta\rho)(\sin(QR) - QR \cos(QR))}{(QR)^3} \right]^2$$

$$I(Q) = \frac{2\phi(\Delta\rho)^2 Zv_m (e^{-x} + x - 1)}{x^2} \quad x = (QR_g)^2$$

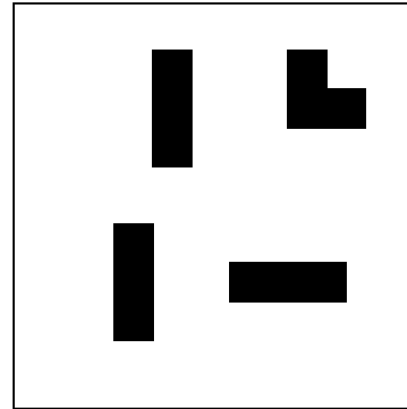
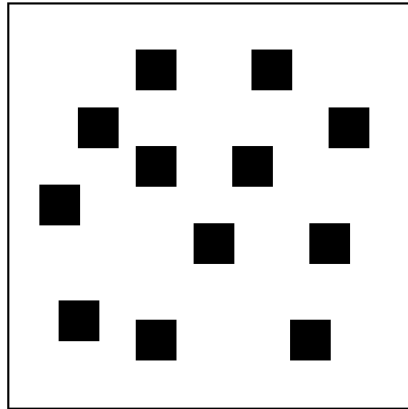


Model Independent Analysis

What information can you obtain?

- Invariant
 - volume fraction, data consistency
- Porod Limit
 - specific surface area, surfactant head group area
- Guinier Analysis
 - general or specific dimensions
- $I(Q=0)$
 - particle volume, molecular weight

Scattering Invariant



10 % black
90 % white
in each square

- Scattered intensity for each would certainly be different

$$Q_I \equiv \int_0^{\infty} q^2 \frac{d\Sigma}{d\Omega}(q) dq$$

- For an incompressible, two-phase system:

$$Q_I = 2\pi^2 \Delta\rho^2 \phi(1 - \phi)$$

- Domains can be in any arrangement

*Guinier and Fournet, pp. 75-81.

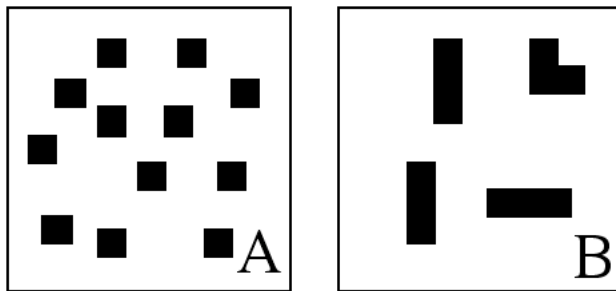
**Need “wide” Q-range to do integration

Porod Scattering

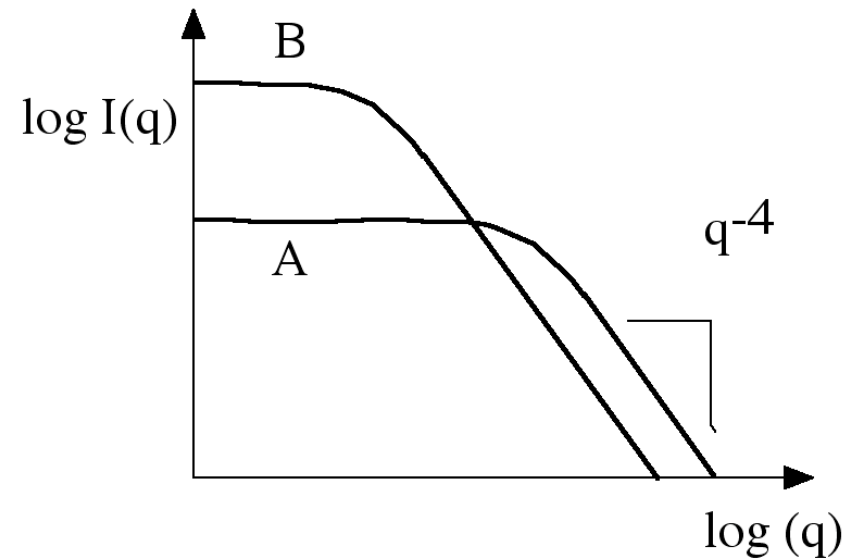
- At large q : $I(q) \propto q^{-4}$

$$\lim_{q \rightarrow \infty} \frac{d\Sigma}{d\Omega}(q) \equiv C_p / q^4 = 2\pi \Delta\rho^2 S_V / q^4$$

S_V = specific surface area of sample



$$\frac{S_A}{V} > \frac{S_b}{V}$$



*Glatter and Kratky, pp. 30-31.

**Need “sharp” interface and “high” Q

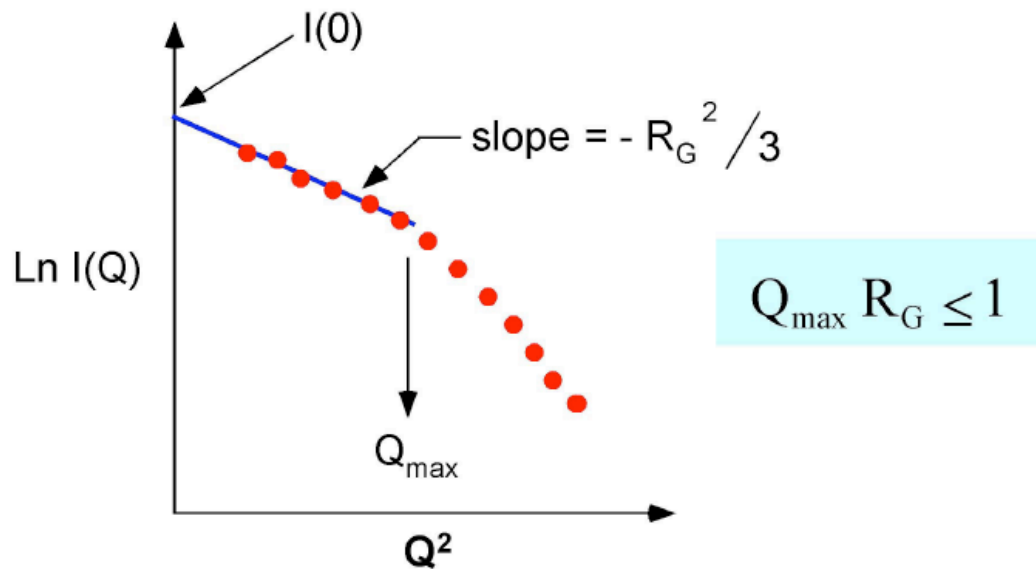
Guinier Analysis

Guinier Approximation:

$$I(Q) \cong I(0)e^{-\frac{1}{3}R_G^2 Q^2}$$

Guinier Plot:

$$\ln[I(Q)] = \ln[I(0)] - Q^2 R_G^2 / 3$$



**Need “dilute” particles and “low” Q

Guinier Radius = R_g
 = RMS distance from “center of scattering density”

Sphere:

$$R_g^2 = \frac{3}{5} R^2$$

Cylinder:

$$R_g^2 = \frac{L^2}{12} + \frac{d^2}{8}$$

Gaussian Coil:

$$R_g^2 = \frac{1}{6} \langle L^2 \rangle$$



Zero angle scattering

$$I(Q = 0) = \frac{1}{V} \left(\int_V \rho(\vec{r}) d\vec{r} \right)^2 \quad \text{becomes} \quad \boxed{I(Q = 0) = \frac{N}{V} (\rho_p - \rho_o)^2 V_p^2}$$

for N uniform particles in volume V, each with SLD ρ_p and volume V_p

In terms of concentration:

$$c(\text{mg/ml}) = \frac{N}{V} \rho V_p$$

$$M_w = \rho V_p N_A \quad \rho = \text{mass density}$$

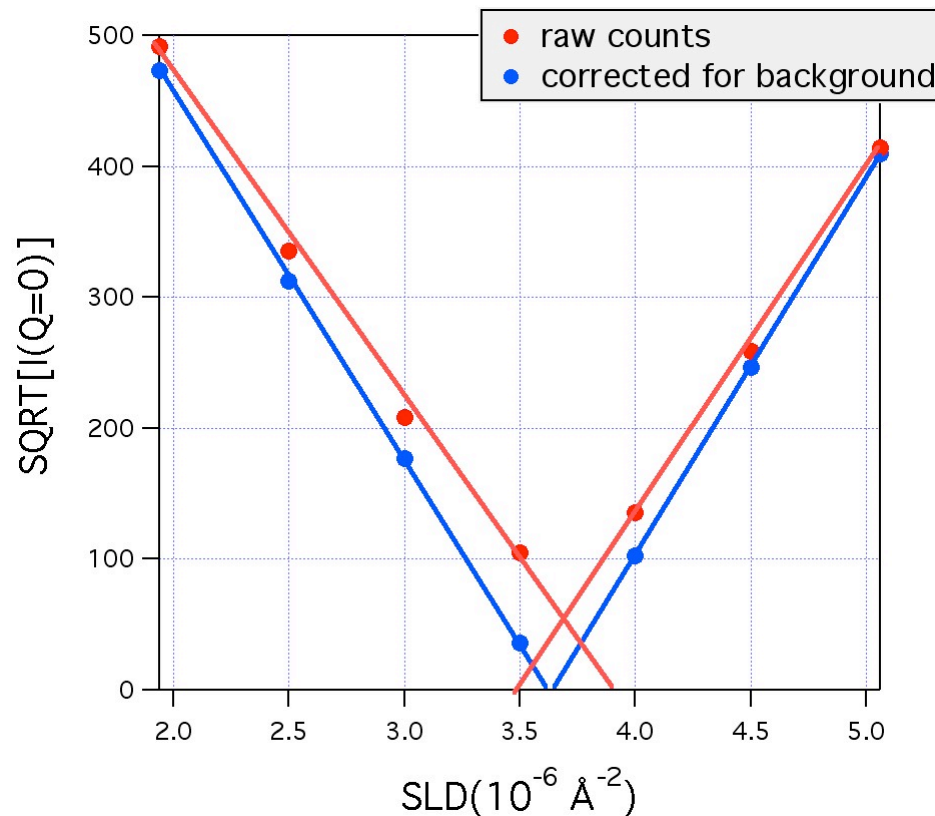
$$\boxed{I(Q = 0) = \frac{c M_w}{\rho N_A} (\rho_p - \rho_o)^2}$$

**Need “dilute” particles and “low” Q



Determining the Contrast Match Point

$$I(Q=0) = \frac{N}{V} (\rho_p - \rho_o)^2 V_p^2$$



- Make several measurements at different solvent SLDs
- Keep the same concentration
- Extrapolate data to $I(Q=0)$
- Plot $\text{sqrt}(I(Q=0))$ vs. SLD

- Don't forget to correct for the incoherent background contribution

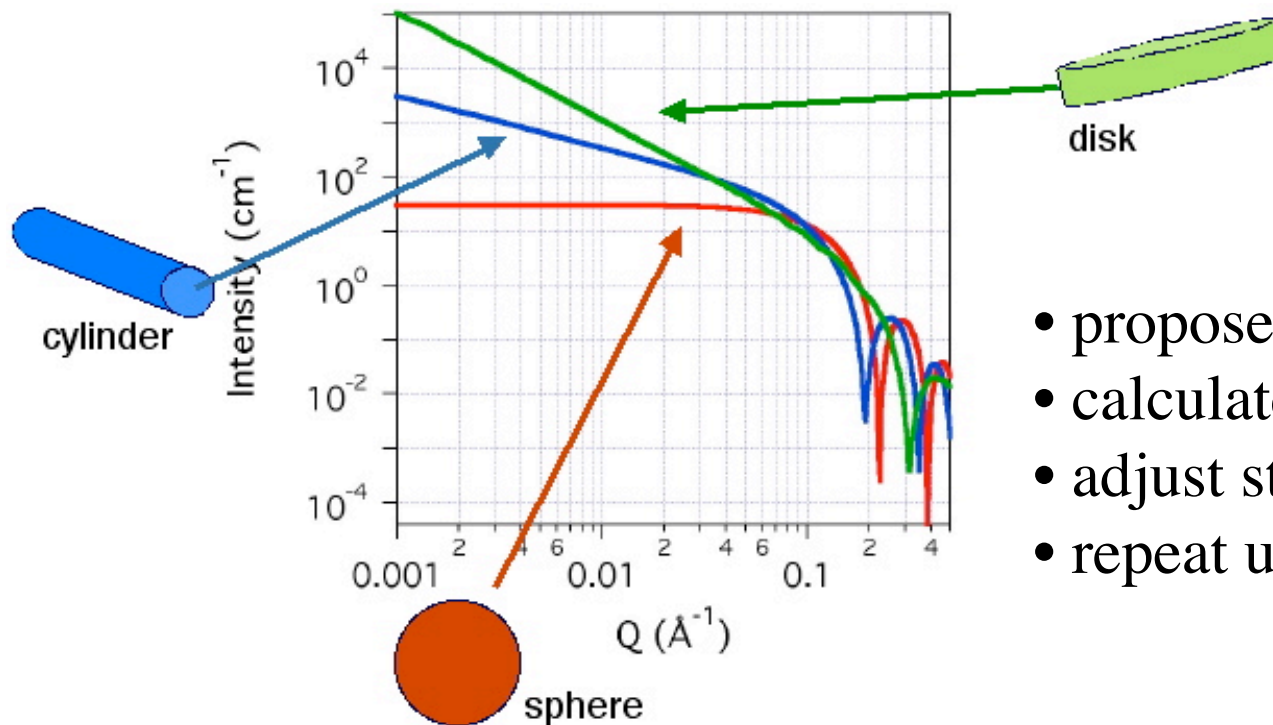
- For composite particles, $I(Q=0)$ will never reach zero - but it will be a minimum at the average particle contrast

**Need "dilute" particles and "low" Q

Non-Linear Model Fitting

One of the most commonly used methods

- a “forward” calculation
- many structures and interactions to choose from

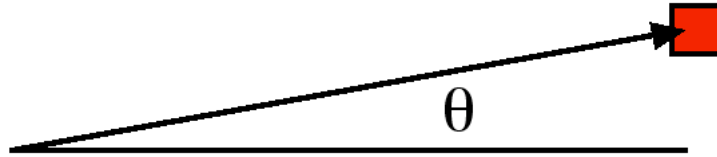


- propose a structural model
- calculate $I(Q)$
- adjust structural parameters
- repeat until done

Resolution

All measured data is affected to some extent by the instrument configuration = “Resolution Smearing”

“True” $I(Q)$ \rightarrow $I_s(Q)$



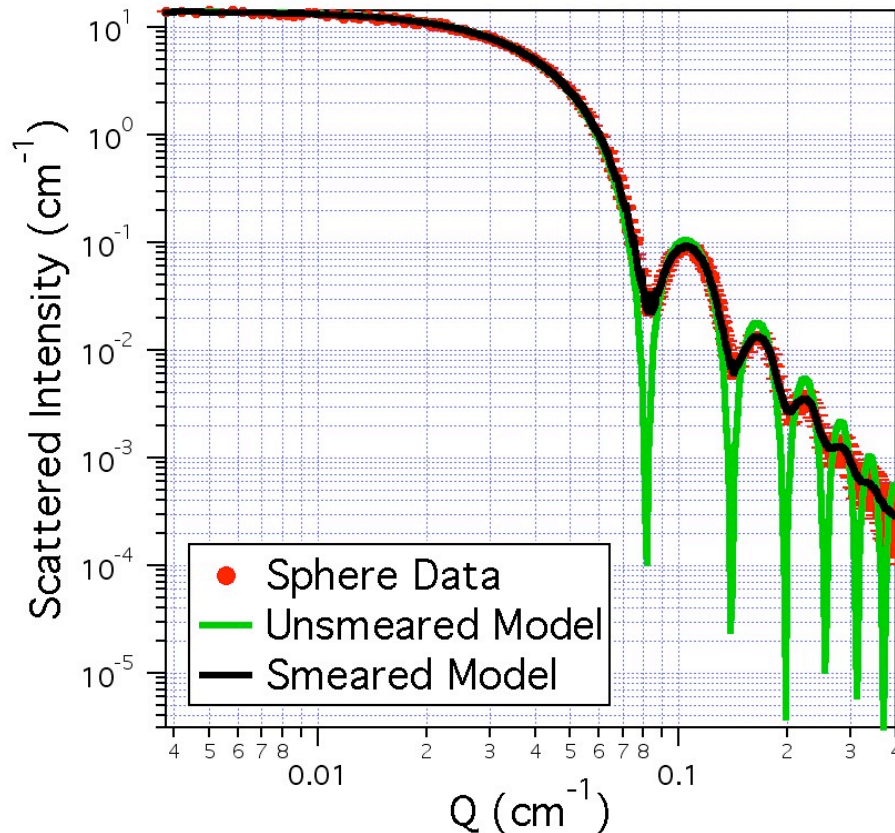
$$Q = \frac{4\pi}{\lambda} \sin(\theta/2)$$

- SANS uses pinhole collimation
- USANS uses slit collimation (more significant smearing)

Resolution effects should NEVER be ignored during analysis

- NCNR tools make it easy to include resolution

Non-Linear Model Fitting



Point	parameters_sf	coef_sf	smear_coef_sf
0	scale	0.05	0.05
1	Radius (A)	55	55
2	contrast (Å ⁻²)	2e-06	2e-06
3	bkgd (cm ⁻¹)	0	0

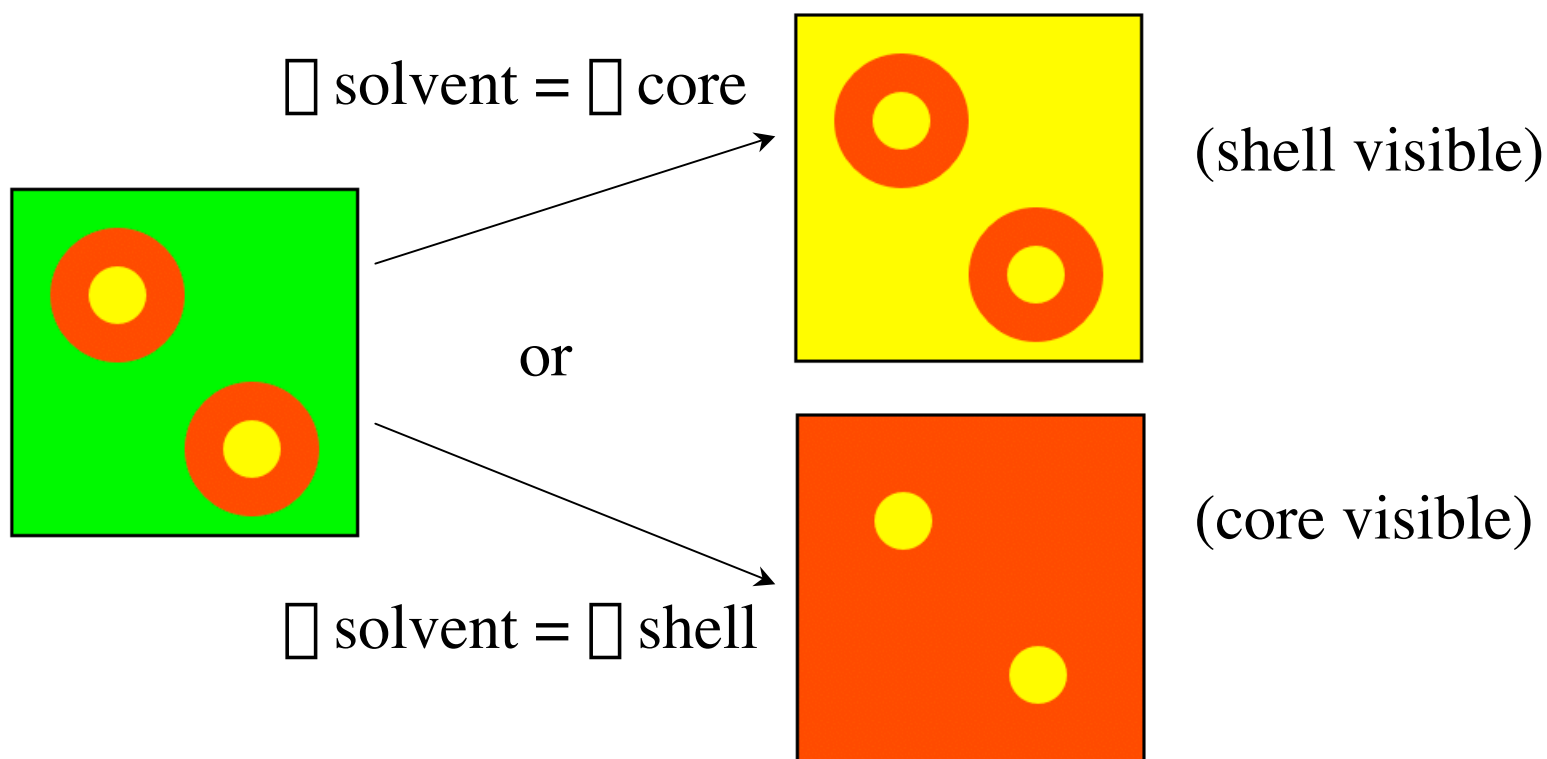
**Need knowledge of sample and model

- Non-linear least squares fitting to experimental data
- Use all the information you can to reduce the number of free model parameters
 - SLD's
 - Concentrations
 - Lengths
- A “good” fit does not necessarily guarantee a perfect representation of the structure in the sample

Contrast Variation

Contrast Matching

reduce the number of phases “visible”

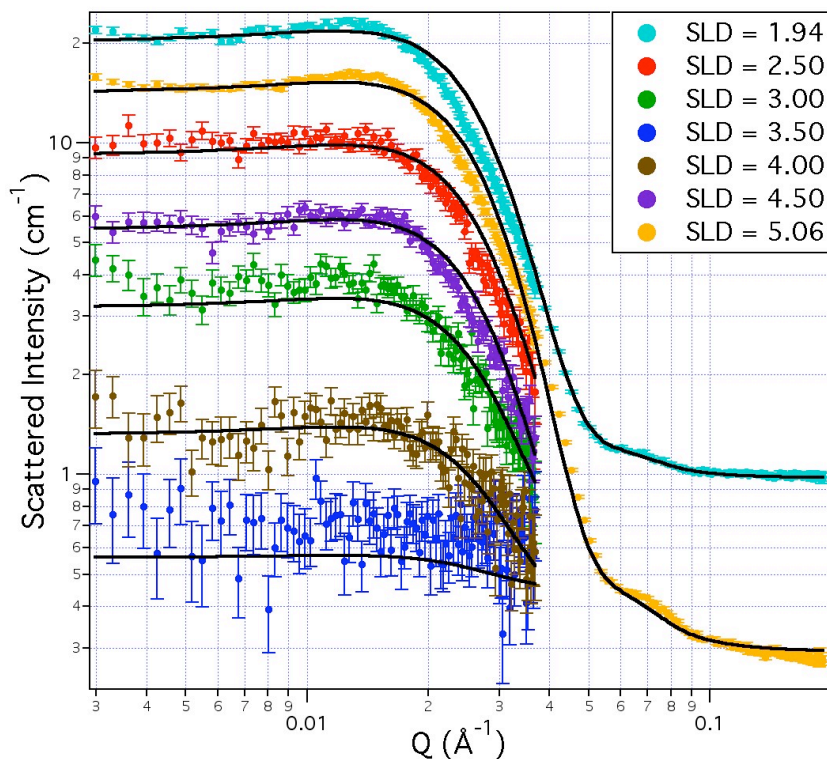


- The two distinct two - phase systems can be easily understood

Global Fitting

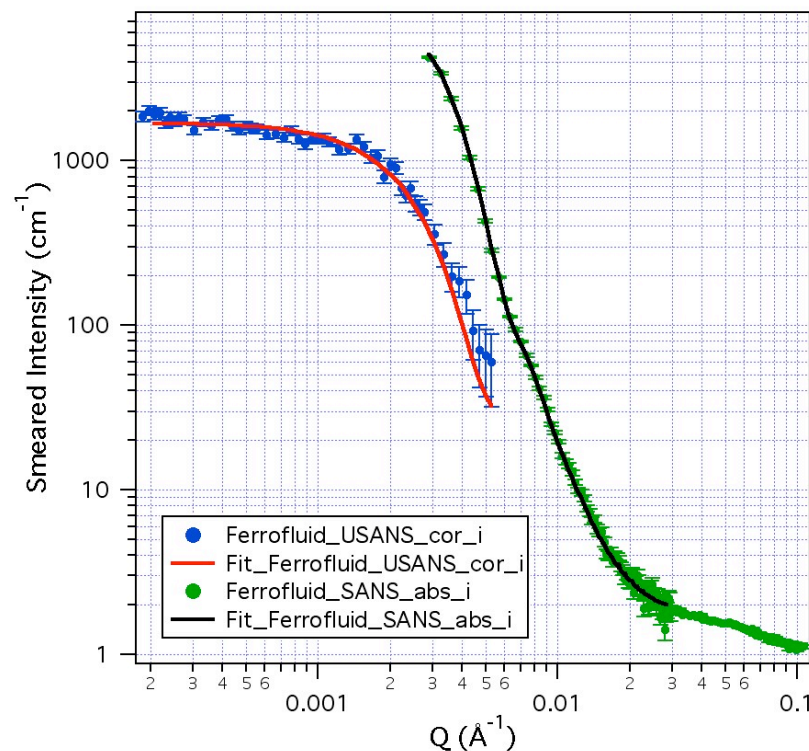
Contrast variation

- same particles, different solvent
- R , ρ_p , ρ are the same
- ρ_{solv} , background are different



SANS + USANS data

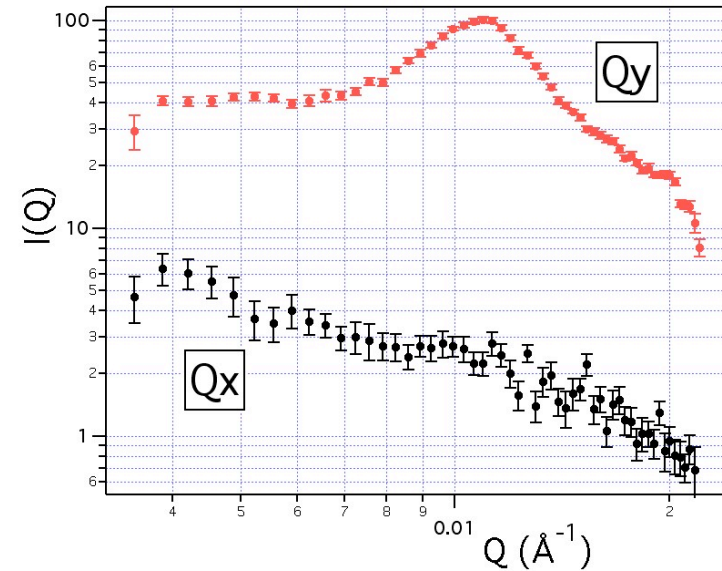
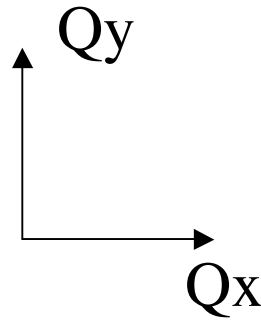
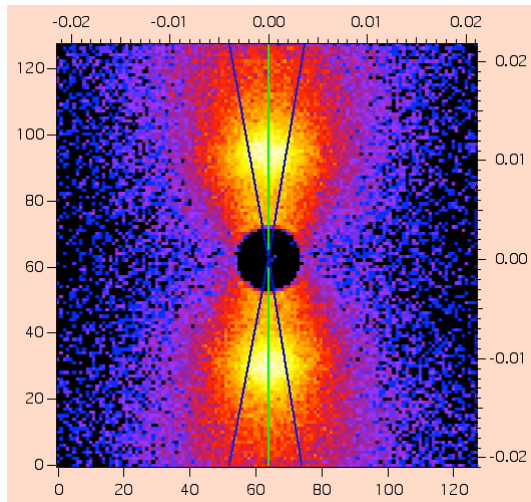
- same sample (same cell)
- all parameters are the same
- smearing, scaling different



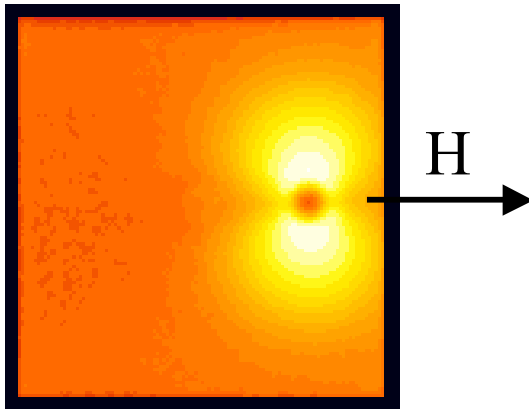
**Need “correct” model

Anisotropic Scattering

Elongated particles aligned by shear



Magnetic domains under an applied field



$$I(Q) \propto (\Delta\rho^2 + \Delta M^2 \sin^2 \phi) P(Q) S(Q)$$

$\square M$ = magnetic contrast

Analyze as a 2D pattern $I(Q, \square)$

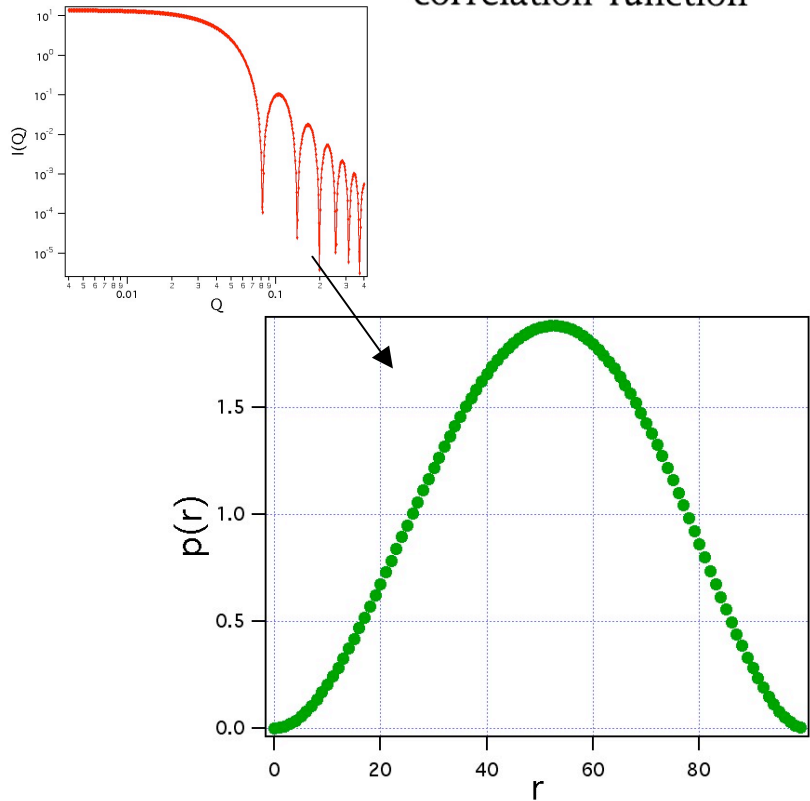
Transforms - $p(r)$

Distance Distribution Function: $p(r)$

$$I_p(Q) \propto \left\langle \int \gamma(\vec{r}) e^{i\vec{Q}\cdot\vec{r}} d\vec{r} \right\rangle = 4\pi \int r^2 \gamma(r) \frac{\sin(Qr)}{Qr} dr$$

↖ average over orientations
↖ correlation function

$p(r)$ is the probability that 2 randomly chosen points are at a distance r apart



If $I(Q)$ is measured over a wide enough Q -range, then one can compute $p(r)$ as the inverse transform:

$$p(r) = \frac{1}{2\pi^2} \int_0^{\infty} I(Q) (Qr) \sin(Qr) dQ$$

For a sphere:

$$p(r) = 12x^2(2 - 3x + x^3) \quad x = r/D$$

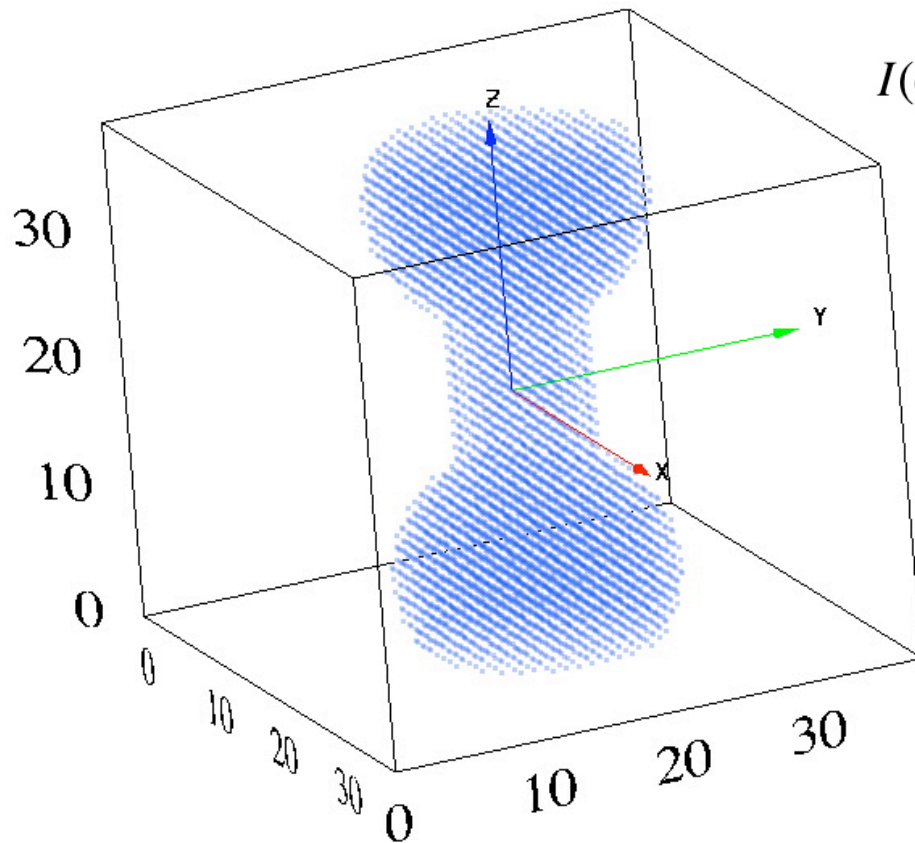
*See: D. Svergun, O. Glatter

**Need “dilute” particles and “wide” Q -range



Ab initio methods

- Calculate $I(Q)$ for complex structures
 - Biological molecules made up of subunits
 - non-standard geometric shapes



$$I(Q) = \sum_{i=1}^n I_i(Q) + 2 \sum_{i \neq j} F_i(Q) F_j(Q) \frac{\sin(Qr_{ij})}{Qr_{ij}}$$

- fill volume with spheres on a grid
 - need distance between every pair
 - can be computationally intensive
 - can optimize shape
- packages available (D. Svergun)

**Need “dilute” particles and “burly” computer

Summary

- Start Simple



- Work up to more complex



- Must use all other information available
- Must always make physical sense
- Must always check that approximations are valid
 - dilute, random, length scales, etc.

- Many tools available at the NCNR
- Many tools available on the web